Journal of Pharmaceutical and Biomedical Analysis

INSTRUCTIONS FOR AUTHORS

Contributions which fulfil the Aims and Scope of the journal will be welcomed from anywhere in the world. The language of the journal is English. All manuscripts should be written in the past tense and impersonal style.

Manuscript Format: Manuscripts should be type-written on good quality paper conforming to either European A4 size (210 × 297 mm) or US Letter size (8.5 × 11 inches). Manuscripts must be *double-spaced on one side only*, with at least 2.5cm (1 inch) margins all round and should be submitted *in triplicate*. All pages should be numbered and the first page must contain the following: title, names of all authors with their addresses in full. The name of the corresponding author should be indicated by an asterisk and added as a footnote to the first page, which should be followed by the following sections in sequence: *Abstract* (Reviews and Full Papers only); *Keywords*; *Introduction*; *Experimental* (or *Materials and Methods*); *Results*; *Discussion*; *Conclusions*; *Acknowledgement(s)*; *References*; *Tables* (each on an individual page with legend); list of *Figure Legends*; *Figures*. Figures and Tables must *not* be included in the body of the text.

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Experimental (or *Materials and Methods*): This section should contain reasonably detailed accounts of materials and experimental procedures, and/or references to previously published methods used. Sufficient information should be provided to permit repetition of the work by other workers. When describing mixed solvents for chromatography or electrophoresis, extraction or other purposes, the following convention must be adopted: solvent A–solvent B–solvent C (a:b:c, v/v/v) or (a:b:c, w/w/w) where a:b:c are the proportions (by volume or weight as appropriate) of the components A, B and C, respectively.

The method of preparation of buffers should be clearly expressed, with the pH value and molarity stated in parentheses, e.g. sodium acetate (pH 4.7; 0.1 M). For mixed solvent systems, it should be clearly stated whether the pH value quoted is the pH of the *original* aqueous component or the *apparent* pH (i.e. pH*) of the mixed solvent system. Typical examples of mobile phases employed in liquid chromatography might be: acetonitrile–sodium octylsulphate (10 mM)–sodium acetate (pH 4.7;0.1 M) (25:25:50, v/v/v), and acetonitrile-sodium octylsulphate (10 mM)–sodium acetate (0.1 M) (25:25:50, v/v/v)(pH* 4.7).

Results: The important results of the work should be clearly stated and illustrated where necessary by tables and figures. The latter should be kept to the minimum consistent with clarity. In particular figures showing linear analytical response curves are generally unnecessary, and will be deleted. The details of slope, intercept, standard error of slope, standard error of intercept, concentration range and number of standards are essential and they should be given in the text or tabulated. This section may also contain experimental detail such as that obtained when describing the development of new analytical procedures. It should include all relevant validation data, e.g. precision and reproducibility at defined concentrations for *n* replicates, limit of quantitation (if appropriate), limit of detection (if appropriate), accuracy, recovery, selectivity, specificity, robustness, ruggedness etc.

Discussion: The results, and their wider implications, should be fully discussed. In some cases, this section may conveniently be combined with the *Results* section.

Conclusions: Where appropriate, a section may be included, which concisely summarizes the principal conclusions of the work and highlights the wider implications. This section should not merely duplicate the abstract.

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References: References cited in the text should give *inclusive page numbers*. They must be definitive citations in the literature. References may include papers that have been accepted for publication and are in press. It is important that each citation should refer to a single work. References must be cited using numerals in brackets, e.g. [23]. Citations in the list of *References* at the end of the paper should be in numerical order of citation using the formats:

[23] F. Maxl, W. Slehr, J. Pharm. Biomed. Anal., 7 (1989) 211–216.

[24] K. Imai, T. Toyooka, in: R.W. Frei, K. Zech (Eds.), Selective Sample Handling and Detection in High-Performance Liquid Chromatography, Part A, Elsevier, Amsterdam, 1988, pp. 209–288.

Complete books should be cited as in [24] above, without the addition of page numbers, and with the editors' or authors' names first. Journal-name abbreviations should be those adopted by the Chemical Abstracts Service (Bibliographic Guide for Editors and Authors, The American Chemical Society, Washington, DC, 1974). If the correct abbreviation is not known, the title should be given in full.

Tables: Should each be typed on a separate page, numbered in sequence with the body of the text. Tables should be headed with a short, descriptive caption. They should be formatted with horizontal lines only: vertical ruled lines are not required. Any annotation to the headings or to the tabulated items must be indicated by a superscript letter and added in sequence at the foot of the table.

List of Figure Legends: A list of Figure Legends must be submitted on a separate sheet to accompany the figures. Each legend must give a concise description of the figure concerned, together with any essential experimental detail not described in the text. In particular, the key to any symbols or distinctive line formats used on the figure must be given.

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ABBREVIATIONS

Ad libitum	ad lib.
Adsorptive stripping voltammetry	ASV
Alternating current	a.c.
Ampere	A
Ångström	Å
Artificial neural network	ANN
Atmosphere	atm
Atmospheric-pressure chemical ionization	APCI
Atomic absorption spectroscopy	AAS
Atomic emission spectroscopy	AES
Atomic weight	at. wt
Audio frequency	a.f.
Biological oxygen demand	BOD
Boiling point	b.p.
Bovine serum albumin	BSA
British thermal unit	B.t.u.
Calorie	cal
Candela	cd
Candle power	c.p.
Capillary electrochromatography	CEC
Capillary electrophoresis	CE
Capillary-zone electrophoresis	CZE
Centimetre	cm
Central nervous system	CNS
Centre of gravity	c.g.
Chemical ionization	CI
Circa	ca
Circular dichroism	CD
Company	Co.
Corporation	Corp.
Correlation coefficient	r
Coulomb	C
Counts per minute	cpm
Counts per second	cps
Cubic centimetre	cm^3
Cubic inch	in^3
Cubic metre	m^3
Curie	CI
Cycles per second	$c\ s^{-1}$
Dalton	Da
Day(s)	day(s)
Debye unit	D
Decibel	dB
Degrees	
Celsius	°C
Centigrade	°C
Fahrenheit	°F
Kelvin	K
Degree (temperature difference)	deg.
Degrees of freedom	df
Differential pulse	DP
Differential scanning calorimetry	DSC
Diode-array detection	DAD
Direct current	d.c.
Disintegrations per minute	dpm
Disintegrations per second	dps
Dyne	dyn
Electromagnetic unit	e.m.u.
Electromagnetic force	e.m.f.
Electron Impact	EI
Electron paramagnetic resonance	EPR
Electron spin resonance	ESR
Electron volt	eV
Electrospray ionization	ESI
Enzyme-linked immunosorbent assay	ELISA
Enzyme-multiplied immunoassay technique	EMIT
Erg(s)	erg(s)
Evaporative light scattering	ELS
Feet, foot	ft
Flame-ionization detection	FID
Flow-injection analysis	FIA
Fluorescence polarization immunoassay	FPIA
Food and Drug Administration	FDA
Foot-candle	ft-c.
Foot-pound	ft-lb
Fourier transform	FT
Freezing point	f.p.
Full scan	FS
Gallon	gal
Gas chromatography	GC
Gas-liquid chromatography	GLC
Gauss	G
Gram	g
Gram-molecule	mol
Graphite furnace	GF
Gravitational acceleration	g
Hanging-mercury-drop-electrode	HMDE
Henry	H
Hertz	HF
High frequency	h.f.
High-performance liquid chromatography	HPLC
High-performance thin-layer chromatography	HPTLC
Horse power	h.p.
Hour(s)	h
Human immunodeficiency virus	HIV
Hydrophobic interaction chromatography	HIC
Inch	in.
Inductively coupled plasma	ICP
Infrared	IR
Intermediate frequency	i.f.
Internal diameter	i.d.
Internal unit	I.U.
International Conference on Harmonization	ICH
Ion exchange chromatography	IEC
Ion pair	IP
Isoelectric focussing	IEF
Isotachophoresis	ITP
Joule	J
Kilocalorie	kcal
Kilocycles per second	kHz
Kilogram	kg
Kilometre	km
Kilovolt	kV
Kilowatt	kW
Kilowatt-hour	kWh
Limit of detection	LOD
Limit of quantitation	LOQ
Litre	l
Liquid chromatography	LC
Logarithm	log
Logarithm (natural)	ln
Lumen	lm
Lux	lx
Magnetomotive force	m.m.f.
Mass spectrometry	MS
Megacycles per second	$Me\ s^{-1}$
Megaelectron volts	MeV
Melting point	m.p.
Metre	m
Micellar electrokinetic chromatography	MEKC
Microgram	μg
Microlitre	μl
Micrometre	μm
Micromolar	μM
Micromole	μmol
Miles per hour	m.p.h.
Millicurie	mCi
Milliequivalent	mEq
Milligram	mg
Millilitre	ml
Millimetre	mm
Millimolar	mM
Millimole	mmol
Millisecond	ms
Milliosmolar	mOsM
Minute(s)	min
Molar concentration	M
Molar weight (relative mobility)	M_r
Month	month
Nanometre	nm
Nanomole	nmol
Near-infrared spectroscopy	NIRS
Negative chemical ionization	NCI
Nuclear Overhauser effect	NOE
Normal concentration	N
Normal phase	NP
Nuclear magnetic resonance	NMR
Ohm	Ω
Outside diameter	o.d.
Overpressured layer chromatography	OPLC
Parsec	pc
Partial least-squares	PLS
Particle induced X-ray emission	PIXE
Phosphate-buffered saline	PBS
Picofarad	pF
Positive chemical ionization	PCI
Picomole	pmol
Polyacrylamide gel electrophoresis	PAGE
Pound(s)	lb
Pounds per square inch (in American or technological works)	$lb\ in^{-2}$
Probability	P
Proton magnetic resonance	^{1}H-NMR
Quality control	QC
Quantitative structure-activity relationship	QSAR
Radian	rad
Radioimmunoassay	RIA
Radio-frequency	r.f.

ABBREVIATIONS

Relative humidity	r.h.	Square metre	m^2	Versus	vs
Relative standar deviation	RSD	Standard deviation	SD	Volt	V
Reversed-phase	RP	Standard error	SE	Volt-ampere	VA
Revolutions per minute	rpm	Standard temperature and pressure	S.T.P.	Volt-coulomb	VC
Root mean square	r.m.s.	Supercritical-fluid chromatography	SFC	Volume	vol.
Second(s)	s	Supercritical-fluid extraction	SFE	Volume by volume	v/v
Scanning-electron microscopy	SEM	Surface plasmon resonance	SPR	Watt	W
Sodium dodecyl sulfate	SDS	Thermospray ionization	TSP	Watt-hour	Wh
Solid-phase extraction	SPE	Thermogravimetric analysis	TGA	Weber	Wb
Solid-phase microextraction	SPME	Thin-layer chromatography	TLC	Weight	wt
Square foot	ft^2	Ultraviolet	UV	Weight by volume	w/v
Square inch	$in.^2$	United States Pharmacopeia	USP	X-ray powder diffraction	XRPD

PREFIXES

Prefixes to the Names of Units

Multiplier	Prefix	Symbol	Multiplier	Prefix	Symbol
10^{-1}	deci	d	10	deca	da
10^{-2}	centi	c	10^2	hecto	h
10^{-3}	milli	m	10^3	kilo	k
10^{-6}	micro	μ	10^6	mega	M
10^{-9}	nano	n	10^9	giga	G
10^{-12}	pico	p	10^{12}	tera	T
10^{-15}	femto	f	10^{15}	peta	P
10^{-18}	atto	a	10^{18}	exa	E

AUTHOR CHECKLIST

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Keywords provided . . . . . ☐

Abstract provided . . . . . ☐

References cited in correct form [1], [2,3], [4–6] . . . . . ☐

Reference list corresponds exactly with numbered citations in text . . . . . ☐

Correct abbreviations for Journals, according to the Chemical Abstracts Service . . . . . ☐

References listed with first and last pages . . . . . ☐

SI units and conventions used wherever appropriate . . . . . ☐

Figures

Top copy in black ink on white paper, A4 size, or glossy prints . . . . . ☐

All graph axes labelled outside frame, using SI units . . . . . ☐

Figure number and Author's name given on back of each sheet . . . . . ☐

All abbreviations fully explained in legends . . . . . ☐

Scale marks for graphs drawn INSIDE frame . . . . . ☐

Graphs need complete frame, not just 2 axes . . . . . ☐

Correct symbols for individual points used on graphs. Symbols adequate size to allow for reduction . . . . . ☐

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